

## ***Varroa* mite evolution: A neglected aspect of worldwide bee collapses?**

Nurit Eliash<sup>1,2</sup>, Alexander Mikheyev<sup>2,3\*</sup>

5 <sup>1</sup>Institute of Plant Protection, Agricultural Research Organization, The Volcani Center,  
Bet Dagan, Israel

<sup>2</sup>Ecology and Evolution Unit, Okinawa Institute of Science and Technology, Onna-son,  
Okinawa, Japan

<sup>3</sup>Research School of Biology, Australian National University, Canberra, ACT, Australia

10 Email: [sasha@homologo.us](mailto:sasha@homologo.us), [norikachan@gmail.com](mailto:norikachan@gmail.com)

### **Highlights:**

- Breeding bees for *Varroa* tolerance has largely ignored potential mite adaptation
- *Varroa* genetic diversity is greater than previously envisioned and they adapt
- 15 rapidly
- We suggest that tolerance is best viewed as a ‘shared trait’ of hosts and parasites
- Focusing on the interaction between partners can inform research and breeding

### **Abstract**

While ectoparasitic *Varroa* mites cause minimal damage to their co-evolved ancestral  
20 host, the eastern honey bee (*Apis cerana*), they devastate their novel host, the western  
honey bee (*Apis mellifera*). The host switch caused worldwide population collapses,  
threatening global food security. *Varroa* management strategies have focused on  
breeding for bees for tolerance. But, can *Varroa* overcome these counter-adaptations in a  
classic coevolutionary arms race? Despite increasing evidence for *Varroa* genetic diversity  
25 and evolvability, this eventuality has largely been neglected. We therefore suggest a  
more holistic paradigm for studying this host-parasite interaction, in which ‘*Varroa*-  
tolerant’ bee traits should be viewed as a shared phenotype resulting from *Varroa* and  
honey bee interaction.

**Keywords:** coevolution, honey bees, host switch, parasites, tolerance

## 30 **Introduction**

Honey bee populations decline in many countries worldwide. This phenomenon has both ecological and economical impacts, as honeybees are the main pollinators in most agricultural systems, as well as in natural habitats (Paudel et al. 2015; Potts et al. 2010; Hung et al. 2018). It is now well established that the reasons for this decline are  
35 multifactorial, but primarily driven by *Varroa* mites and viruses that they vector (Steinhauer et al. 2018). In the colony of its original host, the eastern honey bee (*Apis cerana*), *Varroa* mites are tolerated while causing minimal damage (Rosenkranz et al. 2010). However, due to the globalization of beekeeping, eastern honey bees and their mites have come into contact with the western honey bee (*Apis mellifera*). This caused  
40 exceptionally rapid switches by two mites (*Varroa destructor* and *Varroa jacobsoni*) to this new host, causing colony collapse and damage estimated in billions of dollars (Gallai et al. 2009). As the use of chemical pesticides has been severely reduced in many countries, one of the notable control approaches is to breed for “*Varroa* tolerant” bees (Carreck 2011). This review aims to highlight the mites’ role as actively adapting  
45 members of the coevolutionary interaction with honey bees, an observation that has significant pest control implications.

### **‘*Varroa*-tolerant’ bees and their genetic basis**

Honey bees have experienced diverse selective pressures, but domestication by humans led to a major turning point in their evolution. From the first record of honey bees  
50 domestication in Egypt 2600 BCE, bee colonies were selected for traits beneficial to humans. At first probably this may have been an incidental consequence of beekeeping, but later as a result of sophisticated breeding programs. The main desired traits were high honey yield and gentle temperament, but also tolerance of diseases and pathogens (vanEngelsdorp & Meixner 2010). While bee diseases have been a continuing problem  
55 for centuries, the emergence of the *Varroa* mite as a particularly devastating pest in *A.*

*mellifera* colonies has caused particular concern. Beekeepers as well as bee researchers have bred for traits that may help the bee colony to cope and survive mite infestation, particularly as a long-term and sustainable alternative to pesticides. This approach has solid theoretical justification, given that millions of years of coevolution have allowed the  
60 ancestral host, the eastern honey bee (*Apis cerana*) to tolerate the mite, and some subspecies of *A. mellifera*, such as the African and Africanized varieties are naturally resistant (Mondragón et al. 2006). In addition, several previously susceptible populations have evolved to tolerate *Varroa* (Seeley 2017; Brettell & Martin 2017; Locke 2016; De Jong & Soares 1997; Mikheyev et al. 2015). In that spirit, some breeding programs have  
65 been selecting bee colonies using “live and let die” strategy, in which the colonies remain untreated and only a small portion of the surviving colonies will allow to contribute queens and drones for the next generations (Kefuss et al. 2004). For a few of the ‘*Varroa*-tolerant’ lines the mechanism of *Varroa* tolerance was investigated, and, increasingly, the genetic basis underlying the tolerance is becoming understood, potentially improving the  
70 efficiency of these programs.

The bee breeding programs have resulted in a number of widely known and commercially used lines. In the US the three ‘*Varroa*-tolerant’ lines: the *Varroa* Sensitive Hygiene (VSH), the Russian honey bees, and the Minnesota Hygienic lines (Spivak et al. 2009; Rinderer et al. 2010). In Europe, several tolerant lines were bred by natural  
75 selecting from local lines, such as in France (Kefuss et al. 2015), and in Norweig (Swenson et al. 2018). In addition, *Varroa*-targeted breeding programs have been concentrated by the COLOSS initiative, that leads several pan-European experiments to assess the *Varroa* tolerance capacity of local bees (<http://coloss.org>).

Over the past decade, as molecular methods have improved and became less costly, our  
80 knowledge of bee genetics increased with the growing understanding of the bee genome (reviewed by (Grozinger & Robinson 2015; Niño & Cameron Jasper 2015)). Altogether, these advances enabled the identification of genetic markers, those are aimed to improve breeding for healthier bees, and shorten this years-long process. An obvious great

attention was given for the search of markers for ‘*Varroa*-tolerant’ bees, by trying to  
85 correlate specific markers to specific useful traits (Zakar et al. 2014) (Figure 1). More  
recently, a few studies exploited the approach of genome-wide association study (GWAS)  
to detect specific markers, those include SNPs for bees’ hygienic behavior (Spötter et al.  
2016), mitochondrial DNA SNPs that discriminate between *Varroa* susceptible and  
tolerant bee colonies (Kim et al. 2019), and ecdysone-induced gene in bee pupa that was  
90 found to affect *Varroa* reproduction (Conlon et al. 2019).

### ***Varroa* genetic variance**

While bee genetic architecture is well investigated, little is known about how *Varroa*  
have evolved in the last 60 years since its shift to *A. mellifera*. *Varroa* were generally  
believed to be clonal populations with low genetic variability according to surveys using  
95 randomly amplified polymorphic DNA (RAPD) markers (Kraus & Hunt 1995),  
mitochondrial DNA, and microsatellites (Solignac et al. 2005). *Varroa* colonies regularly  
sib-mate and have correspondingly high inbreeding coefficients (Broeckx et al. 2019).  
However, more recent studies using different sampling regimes (Dietemann et al. 2019;  
Gajić et al. 2019), and whole-genome data (Techer et al. 2019), show *Varroa* genetic is  
100 much more diverse than thought before. Regardless of underlying genetic diversity,  
extensive evidence exists that *Varroa* experience high selective pressures and rapidly  
evolve in response to pesticides treatments. Pyrethroids resistant mites were reported  
across Europe (Martin 2004), the UK (Thompson et al. 2003), and the middle east  
(Israel) (Mozes-Koch et al. 2000). Interestingly, this resistance can be reversed, when  
105 stop exposing for a few years, suggesting a potential cost (Milani & Della Vedova 2002).  
In addition, mites evolved rapid resistance for other chemical families such as  
organophosphorus (Elzen et al. 2002; Spreafico et al. 2001), and Formamide  
(Rodríguez-Dehaibes et al. 2005; Maggi et al. 2010). And in some cases, mites showed  
resistance to multiple active chemicals, which makes impossible to rotate between  
110 available pesticides (Sammataro et al. 2005). Consequently, *Varroa* mites do have a  
potential for rapid evolution, at least vs. chemicals, and deserves further consideration.

### **“*Varroa* tolerant” traits as host-parasite shared traits**

A key property of coevolutionary systems highlighted in the host-parasite literature is the “shared control” of some traits, namely those that emerge from the joint action of the interacting genotypes (Restif & Koella 2003). Classically, antagonistic coevolution can result in arms races centered around key traits such as host resistance/tolerance, and pathogen virulence. However, while this is well-established in other agricultural systems such as plant diseases (Lambrechts et al. 2006; Sacristán & García-Arenal 2008), it has received less attention in the study of invertebrate parasites such as *Varroa* mites.

Although all bee tolerance traits involve direct interaction with *Varroa*, until recently the possible contribution of *Varroa* genetics to these phenotypes was generally overlooked (Figure 1). However, this has been changing recently. Beaurepaire et al. (2019) have noted the ability by *Varroa* to adapt as a possible factor in a host-parasite arms race, as changes in the genetic structure of mite population in ‘*Varroa*-tolerant’ colonies were higher compared to mites in susceptible colonies, a realization that has led to recent integrative work examining the effect of host genetics on the shared phenotype. For example, Broeckx et al (2019) compared reproducing and non-reproducing mites using DNA microsatellites but found no difference. Recent work has also tried to disassociate the bee-*Varroa*-virus complex by examining the virus effect only (Remnant et al. 2019; Thaduri et al. 2019). While this work provides illuminating insights into bee-virus interactions, we would like to caution that an artificial uncoupling between the *Varroa* and bee genomes ignores the possibility that they actually interact, with important consequences for the course of infection. This may lead to failure in detecting important loci that facilitate the *Varroa* and bee adaptation, and misinterpretation of results for breeding programs and research of ‘*Varroa*-tolerant’ bees. This idea can be extended further, since social immunity requires the interaction between bee brood, adult bees and *Varroa*, requiring careful consideration of the contribution of each.

In general, coevolution favors a stable equilibrium between host and parasite (Restif & Koella 2003). The equilibrium point depends on diverse genetic and environmental

140 factors and cannot be predicted or generalized (Read 1994; Thompson 2005; Techer et al. 2019). However, in the original association between *Varroa* and *A. cerana*, *Varroa* virulence is fairly attenuated (Rath 1999; Lin et al. 2018). It is therefore reasonable to hypothesize that this will be the ultimate equilibrium state also for *A. mellifera*. In fact, mathematical modeling suggests that a benign *Varroa* haplotype will outcompete the

145 virulent one (Vetharianiam & Barlow 2006). As a result, it could be that in some cases of reported ‘*Varroa*-tolerant’ *A. mellifera* colonies in the wild, the survival of the bees can be also explained by less virulent mite population. *Varroa* may be evolving to reduce honey bee colony mortality to provide itself with a longer-lasting resource. Therefore, observed naturally occurring resistance may result from bee or *Varroa* evolution.

150 Incorporation of *Varroa* genetics may broaden our understanding of traits that were so far explained by mechanisms and genetics of the bee only. For example, a few studies showed that ‘*Varroa*-tolerant’ bees have better ability to recognize mite-infested cells compared to control bees (Martin et al. 2002; Mondet et al. 2016). However, this could also be explained by the variance in *Varroa* camouflage abilities, or variance in *Varroa* cuticular profile between colonies (Kather et al. 2015; Le Conte et al. 2015), in addition

155 to differences in bee sensitivity to *Varroa* presence. *Varroa* genetic variance can also explain inconsistency in ‘*Varroa*-tolerant’ traits in bee lines, and “unsuccessful” breeding programs (Odemer 2019), or failures to import ‘*Varroa*-tolerant’ lines, as often experienced in breeding programs (Meixner et al. 2015). However, as these cases are

160 likely underreported, they are harder to interpret. In one natural population, Seeley (2007) found that there was no difference in *Varroa* growth rates in apparently resistant feral colonies and sensitive commercial strains. While the population in question has undergone a strong selective event after arrival of *Varroa* (Mikheyev et al. 2015), whether this or milder mite strains result in its apparent persistence remains unclear.

165 Recovery of natural bee-swarms a few years after *Varroa* introduction were recorded in a few instances, and were suggested to be partially due to variance in *Varroa* virulence (Fries et al. 2006; Villa et al. 2008).

## Implications and conclusions

In conclusion, we suggest that the design of bee breeding programs for desired ‘*Varroa*-tolerant’ traits should be addressed from a broader perspective that include both host and parasite genetics, and treated as a shared traits. *Varroa* genetic variance should be studied and included as one of the factors influencing the capacity of bees to tolerate *Varroa* mite, in addition to environmental conditions (Le Conte et al. 2007; Currie & Tahmasbi 2008) and bee nutrition (Alaux et al. 2011; Huang 2012). Understanding *Varroa* genetic architecture and quantifying its possible contribution to ‘*Varroa*-tolerant’ traits should enable improved breeding programs that will account for possible coevolutionary interactions in the future. Given that *Varroa* is a fact of life for *A. mellifera*, evolution of less virulent strains of *Varroa* over time, as suggested by mathematical modeling (Vetharaniam & Barlow 2006) may decrease the severity of their impact. When studying bee-*Varroa* interactions regular genetic monitoring of both players should be conducted before and during breeding programs. Such temporal surveys will give us a clue about *Varroa* population genetic dynamics in response to changes in selective pressures, either through pesticide treatment or ‘*Varroa*-tolerant’ phenotypes. These data can help improve our *Varroa* management, for example by rational pesticide rotation, and directed selection of bee lines according to the current *Varroa* population. In the long term, such genetic monitoring of the mite-bee population will enable us to predict eruptions of mite infestation, or resistance events for novel pesticides before they occur, by modeling. However, all the available genetic tools to study *Varroa* are low-resolution and new methods are urgently needed.

## Acknowledgments

Funding: ASM was supported by a Future Fellowship from the Australian Research Council (FT160100178). The funding body had no involvement in the preparation of the manuscript. We are grateful to Steven Gutierrez for drawing the summary figure, and for Ada Rafaeli for proofreading the final version of the manuscript.

195

## References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

\* of special interest

200 \*\* of outstanding interest

Alaux, C. et al., 2011. Nutrigenomics in honey bees: digital gene expression analysis of pollen's nutritive effects on healthy and varroa-parasitized bees. *BMC genomics*, 12, p.496.

205 Beaurepaire, A.L. et al., 2019. Population genetics of ectoparasitic mites suggest arms race with honeybee hosts. *Scientific reports*, 9(1), p.11355. \*\* Found that changes in mite population genetic structure were more notable in mite-resistant colonies, compared to mite-susceptible colonies.

210 Brettell, L.E. & Martin, S.J., 2017. Oldest Varroa tolerant honey bee population provides insight into the origins of the global decline of honey bees. *Scientific reports*, 7(1), p.45953. \* A unique evidence of surviving *Varroa* infested *A. mellifera* colonies, suggested that the survival of the colonies is due to a combination of low *Varroa* reproductive rate and low, but highly diverse, viral infection.

215 Broeckx, B.J.G. et al., 2019. Honey bee predisposition of resistance to ubiquitous mite infestations. *Scientific reports*, 9(1), p.7794. \* By comparing drones' genetics (whole exomes) and mites' genetics (DNA microsatellites), they concluded that suppressed mite reproduction trait in *Varroa* surviving colonies is based on bee, and not *Varroa* genetic variance.

220 Carreck, N.L., 2011. Breeding honey bees for *Varroa* tolerance. In N. L. Carreck, ed. *Varroa, Still a problem in the 21st century?*. The International Bee Research Association, pp. 63–70.

225 Conlon, B.H. et al., 2019. A gene for resistance to the *Varroa* mite (Acari) in honey bee (*Apis mellifera*) pupae. *Molecular ecology*, p.mec.15080. \*\* Using genome-wide association analysis they found that ecdysone-induced gene in bee pupa correlate with suppressed *Varroa* reproduction. They then suggest that the mite acquires the bee-hormone during feeding on the pupa, In turn, the bee-derived ecdysone inhibits reproduction initiation in the mite.

Currie, R.W. & Tahmasbi, G.H., 2008. The ability of high- and low-grooming lines of honey bees to remove the parasitic mite *Varroa destructor* is affected by environmental conditions. *Canadian journal of zoology*, 86(9), pp.1059–1067.

230 De Jong, D. & Soares, A.E.E., 1997. An isolated population of italian bees that has

survived *Varroa jacobsoni* infestation without treatment for over 12 years. *American Bee Journal*.

235 Dietemann, V. et al., 2019. Population genetics of ectoparasitic mites *Varroa* spp. in Eastern and Western honey bees. *Parasitology*, pp.1–11. \* Investigation of *Varroa* population genetics found multiple infestations of single host colonies by both mite species (*V. destructor* and *V. jacobsonii*).

Elzen, P.J., Westervelt, D. & Others, 2002. Detection of coumaphos resistance in *Varroa destructor* in Florida. *American Bee Journal*, 142(4), pp.291–292.

240 Fries, I., Imdorf, A. & Rosenkranz, P., 2006. Survival of mite infested (*Varroa destructor*) honeybee (*Apis mellifera*) colonies in a Nordic climate. *Apidologie*, 37, pp.564–570.

Gajić, B. et al., 2019. Coexistence of genetically different *Varroa destructor* in *Apis mellifera* colonies. *Experimental & applied acarology*. Available at: <https://doi.org/10.1007/s10493-019-00395-z>.

245 Gallai, N. et al., 2009. Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. *Ecological economics: the journal of the International Society for Ecological Economics*, 68(3), pp.810–821.

Grozinger, C.M. & Robinson, G.E., 2015. The power and promise of applying genomics to honey bee health. *Current Opinion in Insect Science*, 10, pp.124–132.

250 Huang, Z., 2012. Pollen nutrition affects honey bee stress resistance. *Terrestrial arthropod reviews*, 5(2), pp.175–189.

Hung, K.-L.J. et al., 2018. The worldwide importance of honey bees as pollinators in natural habitats. *Proceedings. Biological sciences / The Royal Society*, 285(1870).

255 Kather, R. et al., 2015. Evidence for passive chemical camouflage in the parasitic mite *Varroa destructor*. *Journal of chemical ecology*.

Kefuss, J. et al., 2015. Selection for resistance to *Varroa destructor* under commercial beekeeping conditions. *Journal of apicultural research*, 54(5), pp.563–576.

260 Kefuss, J. et al., 2004. *Varroa* tolerance in France of *intermissa* bees from Tunisia and their naturally mated descendants: 1993-2004. *American Bee Journal*, 144(7), pp.563–568.

Kim, J.S. et al., 2019. Single-nucleotide polymorphism markers in mitochondrial genomes for identifying *Varroa destructor*-resistant and -susceptible strains of *Apis mellifera* (Hymenoptera: Apidae). *Mitochondrial DNA Part A: DNA Mapping, Sequencing, and Analysis*, 30(3), pp.477–489.

- 265 Kraus, B. & Hunt, G., 1995. Differentiation of *Varroa jacobsoni* Oud populations by random amplification of polymorphic DNA (RAPD). *Apidologie*, 26(4), pp.283–290.
- Lambrechts, L., Fellous, S. & Koella, J.C., 2006. Coevolutionary interactions between host and parasite genotypes. *Trends in parasitology*, 22(1), pp.12–16.
- 270 Le Conte, Y. et al., 2007. Honey bee colonies that have survived *Varroa destructor*. *Apidologie*, 38, pp.566–572.
- Le Conte, Y. et al., 2015. *Varroa destructor* changes its cuticular hydrocarbons to mimic new hosts. *Biology letters*, 11(6). \* Found that mite from *A. cerana* can better imitate the hydrocarbon profile of their bee host, compared to mites from *A. mellifera*.
- 275 Lin, Z. et al., 2018. Reproduction of parasitic mites *Varroa destructor* in original and new honeybee hosts. *Ecology and evolution*, 8(4), pp.2135–2145.
- Locke, B., 2016. Natural *Varroa* mite-surviving *Apis mellifera* honeybee populations. *Apidologie*, 47(3), pp.467–482.
- 280 Maggi, M.D. et al., 2010. Resistance phenomena to amitraz from populations of the ectoparasitic mite *Varroa destructor* of Argentina. *Parasitology research*, 107(5), pp.1189–1192.
- Martin, C. et al., 2002. Potential mechanism for detection by *Apis mellifera* of the parasitic mite *Varroa destructor* inside sealed brood cells. *Physiological entomology*, 27, pp.175–188.
- 285 Martin, S.J., 2004. Acaricide (pyrethroid) resistance in *Varroa destructor*. *Bee World*, 85(4), pp.67–69.
- Meixner, M.D., Kryger, P. & Costa, C., 2015. Effects of genotype, environment, and their interactions on honey bee health in Europe. *Current opinion in insect science*, 10, pp.177–184.
- 290 Mikheyev, A.S. et al., 2015. Museum samples reveal rapid evolution by wild honey bees exposed to a novel parasite. *Nature communications*, 6, p.7991. \*\* Following the introduction of *Varroa* mite in feral colonies, a major loss of haplotype diversity was observed while nuclear genetic diversity stayed almost unchanged. These may explain the survival of natural populations to new pathogens/parasites.
- 295 Milani, N. & Della Vedova, G., 2002. Decline in the proportion of mites resistant to fluvalinate in a population of *Varroa destructor* not treated with pyrethroids. *Apidologie*, 33(4), pp.417–422.
- Mondet, F. et al., 2016. Specific cues associated with honey bee social defence against *Varroa destructor* infested brood. *Scientific reports*, 6, p.25444. \*\* Found that the

- 300           hygienic behavior by adult bees is highly specific against *Varroa*-infested brood cells,  
and induced by specific cues emitted by the infested brood.
- Mondragón, L., Martin, S. & Vandame, R., 2006. Mortality of mite offspring: a major  
component of *Varroa destructor* resistance in a population of Africanized bees.  
*Apidologie*, 37(1), pp.67–74.
- 305           Mozes-Koch, R. et al., 2000. First detection in Israel of fluvalinate resistance in the  
*Varroa* mite using bioassay and biochemical methods. *Experimental & applied  
acarology*, 24(1), pp.35–43.
- Niño, E.L. & Cameron Jasper, W., 2015. Improving the future of honey bee breeding  
programs by employing recent scientific advances. *Current Opinion in Insect Science*,  
10, pp.163–169.
- 310           Odemer, R., 2019. Reproductive capacity of *Varroa destructor* in four different honey bee  
subspecies. *Saudi Journal of Biological Sciences*. Available at:  
<http://dx.doi.org/10.1016/j.sjbs.2019.09.002> [Accessed September 11, 2019].
- 315           Paudel, Y.P. et al., 2015. Honey Bees (*Apis mellifera* L.) and Pollination Issues: Current  
status, impacts and potential drivers of decline. *Journal of Agricultural Science*, 7(6),  
pp.93–109.
- Potts, S.G. et al., 2010. Global pollinator declines: trends, impacts and drivers. *Trends in  
ecology & evolution*, 25(6), pp.345–353.
- Rath, W., 1999. Co-adaptation of *Apis cerana* Fabr. and *Varroa jacobsoni* Oud. *Apidologie*,  
30(2-3), pp.97–110.
- 320           Read, A.F., 1994. The evolution of virulence. *Trends in microbiology*, 2(3), pp.73–76.
- Remnant, E.J. et al., 2019. Direct transmission by injection affects competition among  
RNA viruses in honeybees. *Proceedings. Biological sciences / The Royal Society*,  
286(1895), p.20182452.
- 325           Restif, O. & Koella, J.C., 2003. Shared control of epidemiological traits in a  
coevolutionary model of host-parasite interactions. *The American naturalist*, 161(6),  
pp.827–836.
- Rinderer, T.E. et al., 2010. Breeding for resistance to *Varroa destructor* in North  
America. *Apidologie*, 41(3), pp.409–424.
- 330           Rodríguez-Dehaibes, S.R. et al., 2005. Resistance to amitraz and flumethrin in *Varroa  
destructor* populations from Veracruz, Mexico. *Journal of apicultural research*, 44(3),  
pp.124–125.

- Rosenkranz, P., Aumeier, P. ia & Ziegelmann, B., 2010. Biology and control of *Varroa destructor*. *Journal of invertebrate pathology*, 103, pp.96–119.
- 335 Sacristán, S. & García-Arenal, F., 2008. The evolution of virulence and pathogenicity in plant pathogen populations. *Molecular plant pathology*, 9(3), pp.369–384.
- Sammataro, D. et al., 2005. The resistance of *Varroa* mites (Acari: Varroidae) to acaricides and the presence of esterase. *International Journal of Acarology*, 31(1).
- 340 Seeley, T.D., 2007. Honey bees of the Arnot Forest: a population of feral colonies persisting with *Varroa destructor* in the northeastern United States. *Apidologie*, 38, pp.19–29.
- Seeley, T.D., 2017. Life-history traits of wild honey bee colonies living in forests around Ithaca, NY, USA. *Apidologie*, 48(6), pp.743–754. \*Found that naturally surviving colonies maintained traits over 10 years after *Varroa* introduction, that probably enable them to tolerate *Varroa* with no particular fitness cost.
- 345 Solignac, M. et al., 2005. The invasive Korea and Japan types of *Varroa destructor*, ectoparasitic mites of the Western honeybee (*Apis mellifera*), are two partially isolated clones. *Proceedings of the Royal Society B: Biological Sciences*, 272(1561), pp.411–419.
- 350 Spivak, M. et al., 2009. The future of the MN hygienic stock of bees is in good hands! *American Bee Journal*, 149(10), pp.965–967.
- Spötter, A. et al., 2016. Genome-wide association study of a varroa-specific defense behavior in honeybees (*Apis mellifera*). *The Journal of heredity*, 107(3), pp.220–227.
- Spreafico, M. et al., 2001. First detection of strains of *Varroa destructor* resistant to coumaphos. Results of laboratory tests and field trials. *Apidologie*, 32(1), pp.49–55.
- 355 Steinhauer, N. et al., 2018. Drivers of colony losses. *Current Opinion in Insect Science*, 26, pp.142–148.
- Swenson, J., Dahle, B. & Cuypers, J.C., 2018. *Resistance mechanisms towards the ectoparasitic mite Varroa destructor in two naturally-surviving honey bee (Apis mellifera) populations from Scandinavia*. Norwegian University of Life Sciences. Available at:  
360 <https://brage.bibsys.no/xmlui/bitstream/handle/11250/2565356/Cuypers2018.pdf?sequence=1>.
- Techer, M.A. et al., 2019. Genomic analyses of sibling honey bee ectoparasitic mite species show divergent strategies of adaptation. Available at:  
365 <http://dx.doi.org/10.1101/512988>. \*\* Found that different bee parasites (*Varroa* sp., and *Tropilaelaps mercedesae*) have evolved differently, indicating distinct

selective regimes operating on each of the parasites.

- 370 Thaduri, S. et al., 2019. Disentangling host-parasite-pathogen interactions in a varroa-resistant honeybee population reveals virus tolerance as an independent, naturally adapted survival mechanism. *Scientific reports*, 9:6221. Available at: <https://doi.org/10.1038/s41598-019-42741-6>. \*\* Found that 'Varroa-tolerant' colonies showed lower mortality to similar viral load, compared to susceptible colonies, suggesting the higher survival rates of the 'Varroa-tolerant' colonies is due to viral tolerance.
- 375 Thompson, H. et al., 2003. *Varroa destructor* resistance to pyrethroid treatments in the United Kingdom. *Bulletin of insectology*, 56(1), pp.175–181.
- Thompson, J.N., 2005. *The Geographic Mosaic of Coevolution*, University of Chicago Press.
- 380 vanEngelsdorp, D. & Meixner, M.D., 2010. A historical review of managed honey bee populations in Europe and the United States and the factors that may affect them. *Journal of invertebrate pathology*, 103(SUPPL. 1), pp.S80–S95.
- Vetharanim, I. & Barlow, N.D., 2006. Modelling biocontrol of *Varroa destructor* using a benign haplotype as a competitive antagonist. *New Zealand journal of ecology*, 30(1), pp.87–102.
- 385 Villa, J.D. et al., 2008. Changes in Honey Bee (Hymenoptera: Apidae) Colony Swarming and Survival Pre- and Postarrival of *Varroa destructor* (Mesostigmata: Varroidae) in Louisiana. *Annals of the Entomological Society of America*, 101(5), pp.867–871.
- Zakar, E., Jávora, A. & Kusza, S., 2014. Genetic bases of tolerance to *Varroa destructor* in honey bees (*Apis mellifera* L.). *Insectes sociaux*, 61(3), pp.207–215.

390

395 **Figure 1. Mechanisms of *Varroa* tolerance by bees: much known about the bee, little about the mite.** Typically a '*Varroa*-tolerant' colony will possess a few of the following desired traits: (1) hygienic behavior (removal of dead and diseased brood), (2) grooming behavior, (3) suppression of *Varroa* reproduction and (4) short post-capping stage duration. The bee-genetic basis of these traits is well studied (reviewed by (Grozinger & Robinson 2015; Niño & Cameron Jasper 2015; Zakar et al. 2014)). However, although *Varroa* participates in all these interactions, the role of its genetics has been neglected.

